		City	UTM X	UTM Y
ocation	Address	Charleston		4254199
	221 Linda Vista Dr	Nitro	426368	4250859
	008 Washington Ave	Scott Depot	423030	4258916
403 1	32 Ridgeview Dr	Saint Albans	425261	4250518
	10b Winfield Rd	Nitro	427251	4249850
405 1	419 Sattes Cir	Cross Lanes	431646	4252803
406 1	4 Presidio Pointe	Charleston	432704	4251817
407 4	923 Dempsey Dr	Nitro	428294	4251975
	27 Robin Rd	Saint Albans	425125	4249333
	Central Ave	Saint Albans	424683	4249744
	23 Riverview Dr	Saint Albans	425860	4248486
	520 Barrett St		423993	4261986
	466 Blue Lick Rd	Winfield	430090	4250479
413	136 Cross Lanes Dr	Nitro	431749	4252974
414	5009 Tipperary St	Cross Lanes	430560	4257928
415	1278 Poca River Rd N	Poca	431732	4252851
416	5016 Kilarney Way	Charleston	429444	
	45 Janette Dr	Poca		4257937
	199 Skyline Dr	Scott Depot	422552	4251001
	209 Virginia Ave N	Saint Albans	425398	4259508
420	97 Richard Dr	Poca	429421	
	157 Shawnee Est	Winfield	426308	
421	23 Log Gdns	Nitro	426249	
	5272 Claybank Rd	Charleston	433233	
423	450 Poplar Fork Rd	Scott Depot	421440	
424	1108 Crossroads Vlg	Nitro	426237	
425	1108 Crossroads Vig	Charleston	432940	
426	14 Hardwood Dr	Poca	429625	
427	41 Evergreen Cir	Nitro	426274	
428	408 Dupont Ave	Saint Albans	424987	
429	31 1/2 Willis Ln	Nitro	427447	
430	209 Hillside Dr Apt C	Scott Depot	421866	
431	160 Kilgore Rd	Charleston	433263	4254138
432	5208 Monta Vista Dr	Poca	42988	4257807
433	186 Spruce Ln	Cross Lanes	43281	4 4255620
434	5406 Bailey Rd	Saint Albans	42684	6 4248879
435	67 Kanawha Ter	Charleston	43203	4 4254113
436	5206 Nesting Way	Poca	42959	9 4258583
437	2935 Poca River Rd N	Nitro	42980	3 425423
438	212 Brookhaven Dr		43038	2 425231
439	120 Oakland Dr	Cross Lanes	43015	
440	121 Oakmont Dr	Poca	42810	3 424966
441	811 1st Ave S	Nitro	42658	36 425077
442	102 Juniper St	Nitro	4328	
443	- 11 0.1	Charleston	4272	
444	1 108 Hillside Dr	Nitro	4230	
44!	and the plant	Scott Depot	4301	
44		Nitro	4302	
44		Poca	4272	
44		Saint Albans		
44		Scott Depot	4226	
	0 4500 1st Ave Trir 3	Nitro	4282	.1/ 72330V

Table 2 (cont.). UTM coordinates, consistent with the NAD83 Zone 17N datum, for potential interior dust sampling locations shown in Figure 2; addresses for sampling candidates are also given.

ATTACHMENT B

TENCON, INC. SOIL SAMPLING PROTOCOL FOR DIOXIN IN THE NITRO WEST VIRGINIA AND SURROUNDING AREA

1.0 BACKGROUND AND BASIS

Soil samples are to be collected from residential properties located in and around Nitro West Virginia. The purpose of the soil sampling is to representatively define the surfacial extent of dioxin contamination in the soil at selected locations in and around Nitro West Virginia. These locations were developed as a part of an overall sampling plan prepared by Dr. George C. Flowers (the PLAN) and provided to TENCON by Carpenter Environmental Associates in conjunction with The Calwell Group PLLC. Copy of the PLAN is attached.

2.0 LOCATION, QUANTITY AND RANDOMIZATION OF SAMPLING POINTS

An initial evaluation of the sampling locations indicates that the majority of the identified sampling points are within the Saint Albans, Bancroft and Scott Depot quadrants of the USGS Topological maps. These locations are identified by UTM coordinates which have been provided to TENCON as a part of the above mentioned sampling plan. In the field, a hand held GPS (UTM coordinate capable) unit will be used to locate the predefined sampling points. and to re-locate sample sites, if necessary. A total of 100 sample points have been defined in the PLAN. Of these, TENCON is to collect 70 split soil samples.

It is our understanding that sampling points have been randomized as a part of the preparation of the sampling PLAN, but that the order of points given in the PLAN is not necessarily the collection order. This will allow for some economy of labor in the sample collection process. Samples will be split at time of collection for analysis by TENCON into two sample jars. One of the sample jars will be provided to a representative of Monsanto and the other will be sent to lab for analysis as directed by Carpenter Environmental in conjunction with the Calwell Group.

3.0 SAMPLE COLLECTION

One soil sample will be collected from each location (up to 70) specified in the PLAN and split into two containers. One of the split sample containers will be given to the Monsanto representative from each PLAN location sampled.

The following factors may cause a sample location to be deemed uncollectable:

- The presence of any unique topographic or other physical features at the sampling location, such as the presence of a steep hillside or cliff, a lake or pond, concrete or asphalt paving, barbed wire or electrified fences, etc.
- Any specific physical hazard such as the presence of vicious animals, poisonous plants, or unsafe physical location issues, etc.
- Any location where there is no prior agreement by the owner or occupant to allow entry
 or where the owner or occupant refuses entry, or exhibits aggressive or otherwise
 threatening behavior.

The Calwell Group is responsible for coordinating with landowners and occupants and for coordinating the entry access agreements for each of the sampling locations in the PLAN.

The PLAN specifies that the soil is to be collected from the top two centimeters of the soil at the locations specified and analyzed by USEPA method 4435. Based on the PLAN's specified laboratory analytical method (XDS-Calux) and the limit of detection specified by the Calwell Group's toxicologist (10 parts per trillion), the lab has specified a minimum sample size of 1 gram per sample. Due to the split sample, it will be necessary to collect 20 grams of soil in a given sample. This is a physical sample of approximately 5" x 5" to a depth of 2 centimeters of soil.

4.0 SOIL SAMPLING PROTOCOL AT EACH LOCATION

- 4.1. Prior to sampling, assemble the sampling components for the specific site: cleaned metal (chrome or stainless) sampling auger, metal pan for compositing soil sample, non-powdered latex gloves, stainless steel trowel or spoon, two 4 ounce amber jars with Teflon lined lids, labels, chain of custody.
- 4.2. Vegetative cover will be removed to ground level prior to sample collection. Once the site is located, record the UTM coordinates on the sampling log for that sample number. Open that sampling kit. The metal pan will be used as a compositing surface.
- 4.3. Using the soil auger, four soil cores two centimeters deep will be outlined and manually removed to the compositing pan using the metal auger and collection trowel. It is anticipated that four adjacent 2 ½" wide auger holes to a depth of 2 centimeters are necessary at each location to provide a sufficient quantity of soil to collect the minimum of 20 grams (10 grams each jar) for analysis.
- 4.4. Prior to filling the two jars, soil from the multiple auger holes will be composited and homogenized in the foil lined metal pan using the collection trowel. The composited sample will be divided in half and each half placed in a labeled sample jar.
- 4.5. Any soil remaining after all sample jars are filled is placed back into the auger holes at the end of sample collection activities. The hand auger and other site specific supplies are placed in a bag to be decontaminated prior to reuse. Gloves are placed in a bag for disposal.
- 4.6. Prior to leaving the specific sample location, the soil type, unusual characteristics of the sample location and time of collection are recorded as field notes. Soil samples are packed in a secure container and kept under lock prior to transport to the analytical lab for analysis of dioxin.

This procedure is repeated for each sample depth at all sample locations.

The above procedure is in compliance with commonly acceptable soil sampling methods as outlined in EPA SW 846.

5.0 SPECIFIED ANALYTICAL LABORATORY

Soil samples to be sent for dioxin analysis (USEPA Method 4435) to Zenobiotic Detection Systems
1601 East Geer Street, Suite S
Durham, North Carolina 27704

TENCON, INC. PROCEDURE FOR COLLECTION OF SPLIT INDOOR DUST SAMPLES FROM HOUSEHOLD SURFACES FOR DIOXIN ANALYSIS USING AN OMEGA VACUUM CLEANER

1.0 BACKGROUND AND BASIS

Household dust samples are to be collected from residential properties located in and around Nitro West Virginia. The purpose of the dust sampling is to representatively define the surfacial extent of dioxin contamination in the dust inside residences at selected locations in and around Nitro West Virginia. These locations were developed as a part of an overall sampling plan prepared by Dr. George C. Flowers (the PLAN) and provided to TENCON by Carpenter Environmental Associates in conjunction with The Calwell Group PLLC. A copy of the PLAN is attached.

The collection of indoor dust samples requires a systematic and regimented collection and handling protocol. This sampling method defines the fundamental techniques used to obtain representative and defensible indoor dust samples to support an environmental project or study effort. TENCON recognizes that there may be site-specific circumstances that necessitate deviation from the standard protocol described in this sample collection procedure. Any such deviation will be fully documented and such information provided prior to conclusion of the sampling event so allow an alternate site to be selected as needed.

2.0 LOCATION, QUANTITY AND RANDOMIZATION OF SAMPLING POINTS

Initially the sampling locations will be plotted on the appropriate quadrants of the USGS Topological maps. It is anticipated that the majority of the identified sampling points will be within the Saint Albans, Bancroft and Scott Depot quadrants of the USGS Topological maps. These locations will be identified by specific street addresses which are provided to TENCON as a part of the PLAN. A total of 350 possible sample points have been defined in the PLAN. Of these, TENCON is to collect 200 (divided between parties to 100 each) dust samples.

It is our understanding that sampling addresses have been randomized as a part of the preparation of the sampling PLAN, but that the order of addresses given in the PLAN is not necessarily the collection order. This will allow for some economy of labor in the sample collection process.

Two (duplicate) samples will be collected for analysis by TENCON at each location. One of these samples cassettes will be sealed, labeled and provided to a representative of Monsanto and the other will be sealed, labeled and sent to lab for analysis as directed by Carpenter Environmental in conjunction with the Calwell Group.

3.0 METHOD SUMMARY

Sample collection uses a vacuum with a nozzle/cartridge collection device attached to the end of the vacuum hose. This nozzle is equipped with an integral cartridge that acts as the actual collection surface. The procedure focuses on maximization of sample mass and collection of unbiased split samples, while preventing any cross-contamination through the use of a new nozzle/cartridge system for each sample location.

The vacuum selected is the Omega Vacuum by Atrix Technologies. This vacuum is equipped with a high-efficiency particulate air (HEPA) filter. The vacuum will be used to collect sample mass in the duplicate nozzle/cartridge assemblies from representative areas containing settled dust throughout the study homes. Surfaces to be sampled include tops of curtains, window ledges, door jambs, ceiling fans and air filters.

Recoverable sample material includes dust and particulate matter having cross-sectional dimensions of approximately 0.3 micrometers (µm) and larger.

Samples will be simultaneously collected in two nozzle/cartridge assemblies. Each cartridge is lined with a permeable collection bag capable of capturing particulate matter having cross-sectional dimensions of approximately 0.3 micrometers (µm) and larger. The two cartridges will be fitted to the end of a T-type splitter which is attached to the Omega Vacuum. The cartridges will be used for simultaneous collection of the two samples. Nozzle/cartridge assemblies are then removed, labeled, sealed in foil in a non conductive bag, and stored in a secure shipping container.

Primary sample areas will target living areas in which residents are most likely to spend proportionately longer amounts of time, i.e. family rooms, bedrooms, etc. Tops of ceiling fan blades, tops of draperies, tops of windows and doors, tops of pictures, and window ledges will also be considered primary sample locations because of their propensity to capture dust.

Because the target detection limit is relatively low (10 parts per trillion), the target mass for each cartridge will be 1.0 gram. Consequently, in instances where study homes are relatively small or when extracted dust mass from primary target living areas is not likely to be sufficient to support the detection limit, secondary target areas including hallways and/or additional bedrooms may be systematically sampled to increase the total mass of the dust sample acquired. In extreme cases, it may be necessary to vacate the study home and consider an alternative study home.

At the end of the sampling event, the result will be two sample masses collected in two unique nozzle/cartridge assemblies. One sample nozzle/cartridge will be sealed, stored in foil in a non-conductive plastic bag, logged in the field log, placed in a controlled container, and subsequently provided to a representative of the Monsanto party. The second sample cartridge will be sealed, stored in foil in a non-conductive plastic bag, logged in the field log, placed in a controlled container and subsequently shipped to Xenobiotic Detection Systems in North Carolina, USA for analysis utilizing EPA Method 4435 with a minimum target detection level of 10 parts per trillion (ppt).

4.0 EQUIPMENT/MATERIALS REQUIRED

The following materials will be required to conduct the indoor dust samples. Both disposable and reusable sampling equipment is required.

- 4.1 Omega Vacuum by Atrix Technologies (vacuum cleaner)
- 4.2 Omega Vacuum Six Foot Flexible Hose
- 4.3 T-Type PVC Vacuum Hose Splitter Attachment
- 4.4 Unique, pre-tared Nozzle Attachment with Semi-Permeable Collection Cartridge
- 4.5 Non powdered latex Gloves
- 4.6 Gram Scale (minimum 50 x .01g)
- 4.7 Labels and ballpoint pen or pencil (NO MARKER)
- 4.8 Teflon Tape
- 4.9 Static-Shielding recloseable bags
- 4.10 Methanol
- 4.11 Distilled water

5.0 SAMPLE PROCEDURES

- 5.1 Review field data sheets, and documentation materials to confirm adequate and appropriate field supplies, and sample locations.
- Upon arrival at the sampling location, document field conditions per the field log forms and record all requested general information. Don appropriate field sampling apparel. Take extra latex gloves in the house for subsequent use. (Observe study home for apparent level of indoor settled dust and overall cleanliness. If it appears that study home will not provide an adequate sample mass, it may be prudent to vacate the sample home and seek an alternative location.)
- 5.3 Survey primary and secondary target sample areas and take note in the field log.

At a minimum, primary sample areas will target living areas in which residents are most likely to spend proportionately longer amounts of time, i.e. in a family room, bedrooms, etc. In addition, air filters, tops of ceiling fan blades, draperies, and window ledges will also be considered primary sample target locations because of the propensity to capture settling dust inside the home. Record additional information required on the field log forms, as appropriate.

- 5.4 Assemble the Omega Vacuum
 - 5.4.1 Ensure that the HEPA filter and catch basin are securely in place
 - 5.4.2 Attach the vacuum hose to the vacuum unit and check fit for tightness
 - 5.4.3 Attach the T-type splitter to the vacuum hose and check fit for tightness

- 5.4.4 Weigh two plastic dust collection cartridges independently and note the weight in tenths of a gram in the field log
- 5.4.5 Mark the top and bottom cartridge plugs on the two plastic dust collection cartridges appropriately such that they might be replaced in their original position once the sampling has been completed.
- 5.4.6 Attach a plastic dust collection cartridge to each end of the T-type splitter and check fit for tightness.
- 5.5 The intent is to collect representative dust samples from a given surface by leading first with the left collection nozzle and then with the right collection nozzle. Identify the type of sample area and begin sample collection per the following guidance:
 - 5.5.1 Door frames, tops of windows, tops of curtain areas start at the farthest end of the left side and move towards the center of the "ledge". Then start at the farthest end of the right side and move towards the center of the ledge. Repeat as needed.
 - 5.5.2 Tops of shelves start at the back left corner of the left side and move towards the center of the shelf. Then start at the back right corner of the shelf and move towards the center. Advance the width of the nozzle and repeat as needed.
 - 5.5.3 Fronts of draperies or cabinets. Move the vacuum head slowly and deliberately down in strips the width of the two vacuum heads, then shift to the left or right side to catch the next adjacent surface. Pass the vacuum head down over each strip until it is believed that the sample area is exhausted.
 - 5.5.4 Prior to beginning any new surface, lift the vacuum heads and turn off the vacuum in order to minimize the risk of biasing the split-samples.
 - 5.5.5 TENCON recognizes that there may be site-specific circumstances that necessitate deviation from the standard protocol described in this sample collection procedure. In such case, documentation and record-keeping diligence is of utmost importance.

Continue this process until both plastic dust collection cartridges have collected a visually estimated 1.0 grams of sample mass.

- 5.6 Remove the nozzle from the T-type splitter. Visually inspect the plastic dust collection nozzle/cartridge for integrity. Replace the inlet plug and weigh the nozzle/cartridge assembly. Record the weight on the label.
- 5.7 Replace the bottom plug in the nozzle assembly.
- 5.8 Wrap each cartridge with Teflon tape such that each plug is held firmly in place

- 5.9 Complete the label on each cartridge containing the sample ID#, the start time, the end time, etc. Record this information in the field log along with the final net weight of the nozzle assembly in tenths of a gram.
- 5.10 Place each cartridge in foil and then in a static-shielding baggie and seal. Log any final the observations in the field log.
- 5.11 Provide one of the cartridges to the representative from Monsanto and have them initial the field log, acknowledging receipt of the split-sample.
- 5.11 The remaining cartridge will be shipped to Xenobiotic Detection Systems (XDS) in North Carolina, USA for analysis utilizing EPA Method 4435 with a minimum target detection level of 10 parts per trillion (ppt).

6.0 CONTINGENCY SAMPLE MEASURES

No contingency sample protocol is in place. In the event that an adequate sample is not reasonably collectable, the study house is to be vacated and an alternative study house will be selected.

7.0 OMEGA VACUUM DECONTAMINATION

This sample collection protocol utilizes a vacuum as a sample collection unit with the sampling medium located at the inlet of the vacuum train. Because of this, any contamination collected in the vacuum hose, the vacuum unit, or the unit's HEPA filter will not contaminate the sampling medium. Also, the HEPA filter will prevent any material that is in the vacuum from becoming airborne in subsequent sampling locations. HEPA filters will be inspected nightly as a part of clean up and prep for the next days sampling.

Decontamination of the interior of the sampling equipment prior to each sampling event would constitute an unnecessary time expense and is not recommended as part of this protocol. Units will be surface wiped after to leaving the home and prior to starting the next sampling procedure.

8.0 QUALITY ASSURANCE/QUALITY CONTROL

There are no specific quality assurance activities which apply to the implementation of these procedures. However, the following general QA procedures apply:

- 1. All data and observations must be documented in the field logbooks.
- 2. All instruments must be operated in accordance with operating instructions as supplied by the manufacturer, unless otherwise specified in the work plan.

9.0 DATA VALIDATION

For every 20 samples collected, a second set of duplicate samples will be collected in that home to serve as a lab duplicate if desired. In addition, a nozzle/cartridge assembly field blank will be submitted for every 100 samples.

10.0 HEALTH AND SAFETY PRECAUTIONS

Field crews reserve the right to refuse to sample a location due to concern for their physical well being due to animals, or threatening behavior of the occupants, if encountered.

TENCON's Health and Safety plan as required by the PLAN will be distributed prior to sampling and must be observed and implemented prior to dust sample collection. Chemical exposures are not anticipated, and physical or mechanical hazards are only those that would be found in any typical household environment.

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1.0 BACKGROUND AND BASIS

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2.0 LOCATION, QUANTITY AND RANDOMIZATION OF SAMPLING POINTS

Initially the sampling locations will be plotted on the appropriate quadrants of the USGS Topological maps. It is anticipated that the majority of the identified sampling points will be within the Saint Albans, Bancroft and Scott Depot quadrants of the USGS Topological maps. These locations will be identified by specific street addresses which are provided to TENCON as a part of the PLAN. A total of 350 possible sample points have been defined in the PLAN. Of these, TENCON is to collect 200 (divided between parties to 100 each) dust samples.

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Recoverable sample material includes dust and particulate matter having cross-sectional dimensions of approximately 0.3 micrometers (µm) and larger.

Samples will be simultaneously collected in two nozzle/cartridge assemblies. Each cartridge is lined with a permeable collection bag capable of capturing particulate matter having cross-sectional dimensions of approximately 0.3 micrometers (µm) and larger. The two cartridges will be fitted to the end of a T-type splitter which is attached to the Omega Vacuum. The cartridges will be used for simultaneous collection of the two samples. Nozzle/cartridge assemblies are then removed, labeled, sealed in foil in a non conductive bag, and stored in a secure shipping container.

Primary sample areas will target living areas in which residents are most likely to spend proportionately longer amounts of time, i.e. family rooms, bedrooms, etc. Tops of ceiling fan blades, tops of draperies, tops of windows and doors, tops of pictures, and window ledges will also be considered primary sample locations because of their propensity to capture dust.

Because the target detection limit is relatively low (10 parts per trillion), the target mass for each cartridge will be 1.0 gram. Consequently, in instances where study homes are relatively small or when extracted dust mass from primary target living areas is not likely to be sufficient to support the detection limit, secondary target areas including hallways and/or additional bedrooms may be systematically sampled to increase the total mass of the dust sample acquired. In extreme cases, it may be necessary to vacate the study home and consider an alternative study home.

At the end of the sampling event, the result will be two sample masses collected in two unique nozzle/cartridge assemblies. One sample nozzle/cartridge will be sealed, stored in foil in a non-conductive plastic bag, logged in the field log, placed in a controlled container, and subsequently provided to a representative of the Monsanto party. The second sample cartridge will be sealed, stored in foil in a non-conductive plastic bag, logged in the field log, placed in a controlled container and subsequently shipped to Xenobiotic Detection Systems in North Carolina, USA for analysis utilizing EPA Method 4435 with a minimum target detection level of 10 parts per trillion (ppt).

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5.0 SAMPLE PROCEDURES

- 5.1 Review field data sheets, and documentation materials to confirm adequate and appropriate field supplies, and sample locations.
- 5.2 Upon arrival at the sampling location, document field conditions per the field log forms and record all requested general information. Don appropriate field sampling apparel. Take extra latex gloves in the house for subsequent use. (Observe study home for apparent level of indoor settled dust and overall cleanliness. If it appears that study home will not provide an adequate sample mass, it may be prudent to vacate the sample home and seek an alternative location.)
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- 5.4 Assemble the Omega Vacuum
 - 5.4.1 Ensure that the HEPA filter and catch basin are securely in place
 - 5.4.2 Attach the vacuum hose to the vacuum unit and check fit for tightness
 - 5.4.3 Attach the T-type splitter to the vacuum hose and check fit for tightness

- 5.4.4 Weigh two plastic dust collection cartridges independently and note the weight in tenths of a gram in the field log
- 5.4.5 Mark the top and bottom cartridge plugs on the two plastic dust collection cartridges appropriately such that they might be replaced in their original position once the sampling has been completed.
- 5.4.6 Attach a plastic dust collection cartridge to each end of the T-type splitter and check fit for tightness.
- 5.5 The intent is to collect representative dust samples from a given surface by leading first with the left collection nozzle and then with the right collection nozzle. Identify the type of sample area and begin sample collection per the following guidance:
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 - 5.5.5 TENCON recognizes that there may be site-specific circumstances that necessitate deviation from the standard protocol described in this sample collection procedure. In such case, documentation and record-keeping diligence is of utmost importance.

Continue this process until both plastic dust collection cartridges have collected a visually estimated 1.0 grams of sample mass.

- 5.6 Remove the nozzle from the T-type splitter. Visually inspect the plastic dust collection nozzle/cartridge for integrity. Replace the inlet plug and weigh the nozzle/cartridge assembly. Record the weight on the label.
- 5.7 Replace the bottom plug in the nozzle assembly.
- 5.8 Wrap each cartridge with Teflon tape such that each plug is held firmly in place

- 5.9 Complete the label on each cartridge containing the sample ID#, the start time, the end time, etc. Record this information in the field log along with the final net weight of the nozzle assembly in tenths of a gram.
- 5.10 Place each cartridge in foil and then in a static-shielding baggie and seal. Log any final the observations in the field log.
- 5.11 Provide one of the cartridges to the representative from Monsanto and have them initial the field log, acknowledging receipt of the split-sample.
- 5.11 The remaining cartridge will be shipped to Xenobiotic Detection Systems (XDS) in North Carolina, USA for analysis utilizing EPA Method 4435 with a minimum target detection level of 10 parts per trillion (ppt).

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Sampling Report for the Monsanto Dioxin Site in Nitro, West Virginia

Prepared For:

The Calwell Practice, PLLC

Ву

George C. Flowers, Ph.D.

July 14, 2009



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Sampling Report for the Monsanto Dioxin Site in Nitro, West Virginia

Introduction

A sampling plan, Sampling Plan for the Monsanto Dioxin Site in Nitro, West Virginia dated May 11, 2009, (Flowers, 2009), was generated at the request of Stuart Calwell, Esq., of the Calwell Practice, PLLC. A total of 99 living-area dust samples and 77 soil samples were collected within the Class Area as defined in West Virginia County Civil Action No. 04-C-465, Virdie Allen et al. v. Monsanto Company, et al. In the case of soil sampling stations, the Sampling Design Tool for ArcGIS developed by the NOAA Center for Coastal Monitoring and Assessment (CCMA) was used to generate NAD83 UTM Zone 17N coordinates for potential sampling stations using a simplified, random design (Flowers, 2009). In the event that the generated soil sample locations were inaccessible due to physical/geographical conditions or on private property, soil samples were taken along the roadway in the vicinity of selected stations. Stations were chosen in order to provide good geographic coverage of the Class Area. For dust samples in residences, potential sampling stations were chosen by first selecting residences within the boundary of Zone 2, in the Class Area as defined by Dr. Kirk Brown in his 2008 affidavit submitted in the case cited above (Brown, 2008). A subset of 350 residences distributed within Zones 2, 3, and 4 was drawn from this selection. There were limitations in sampling homes due to owners not being interested or willing to participate in the study. As a result, additional contacts were made with home owners in the Class Area to obtain the requisite number of samples.

Sampling Campaign

Soil Samples

Approximately twenty grams of sample from the top 2 cm of soil were collected from 77 locations in the Class Area as shown on Figure 1 at the coordinates provided in Table 1. As required by field conditions, sample locations were moved and substitute locations sampled as described in the Sampling Plan using the protocols developed by Tencon, Inc. Soils were placed in laboratory supplied glass vials with PTFE/Teflon-lined caps. Samples were collected in accordance with United States Environmental Protection Agency and West Virginia Department of Environmental Protection protocols and procedures. Ten grams of the sampled soil were made available to the Defendant as split samples. All samples were submitted to Xenobiotic Detection Systems (XDS) in Durham, North Carolina, to be analyzed by EPA Method 4435 for dioxin and PCB TEQ.

Dust Samples

Bulk dust samples from living areas of 99 houses in the Class Area were collected in accordance with protocols developed by Tencon, Inc. and the Sampling Plan. Sampling locations are given in Figs. 2-4 and Table 2. Living-area bulk dust samples were collected using a handheld vacuum

sampler with a filter cartridge at the suction tip. Sample collection focused on accumulated dust on heating, ventilation, and air conditioning filters, fan blades, door frames, other elevated surfaces, and other areas where dust may accumulate. These surfaces represent areas where suspended particles would be deposited from ambient living area air. Attempts were made to collect at least 2 grams of sample to provide sample splits to the Defendant's on-site representatives. Although the majority of homes fall into Brown's Zones 2, 3, and 4, two samples were taken outside these zones but within the Class Area. Dust samples were submitted to XDS for analysis by EPA Method 4435 for dioxin and PCB TEQ. In addition, blank cartridges were also submitted for analysis as quality control.

Sample Analysis

Soil and dust samples were submitted to Xenobiotic Detection Systems located in Durham, North Carolina to determine dioxin and PCB TEQs using EPA Method 4435, "Method for Toxic Equivalents (TEQs) Determinations for Dioxin-Like Chemical Activity with the CALUX ® Bioassay." Method 4435 sample processing procedures and use of an affinity column will be employed so that polychlorinated biphenyls (PCBs) will be separated from the chlorinated dioxins and dibenzofurans to determine the TEQ associated with dioxin. Chain of custody forms for all samples acquired during the sampling campaign are reproduced in the Appendix.

References

Brown, Kirk W., 2008, Affidavit in the matter of Virdie Allen et al. versus Monsanto Company, et al.: Putnam County Civil Action No. 04-C-465, Figure 5, p. 14.

Flowers, George C., 2009, Sampling Plan for the Monsanto Dioxin Site in Nitro, West Virginia:

A report prepared for the Calwell Practice, PLLC, 35 p.

National Oceanic and Atmospheric Administration, Sampling Tool for ArcGIS,

http://ccmaserver.nos.noaa.gov/products/biogeography/sampling/welcome.html

ATTACHMENT A

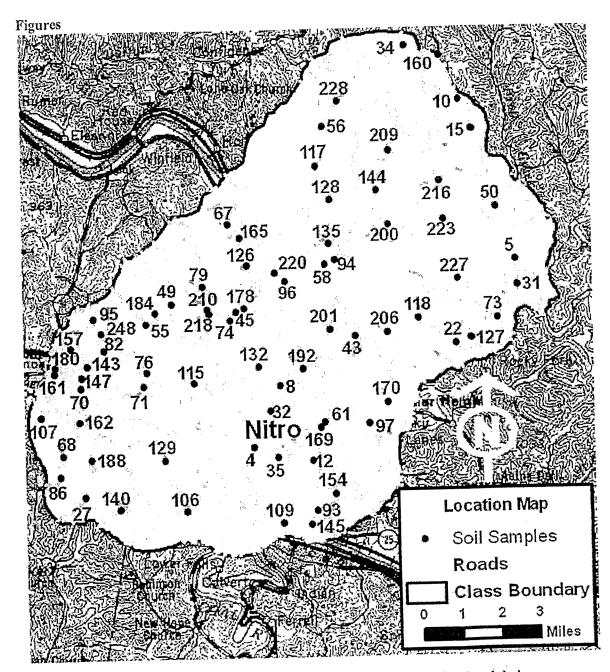


Figure 1. Location map for soil samples taken in the Class Area. Station labels correspond to sample identifications given in Table 1.

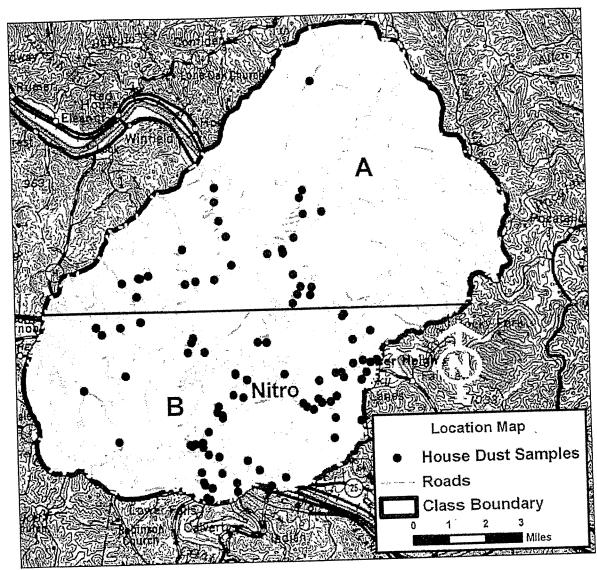


Figure 2. Location map showing interior dust samples taken in the Class Area. Portions of the map indicates by areas A and B are enlarged with station numbers in Figs. 3 and 4.

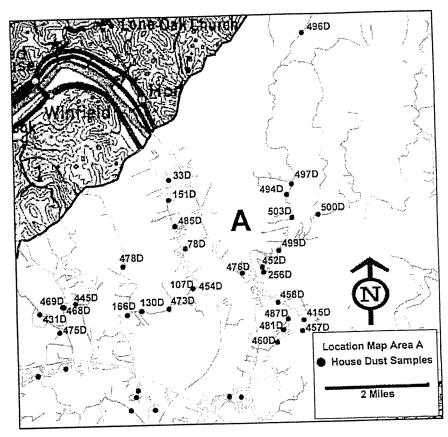


Figure 3. Sample stations for interior dust samples in Area A as shown in Fig. 2.

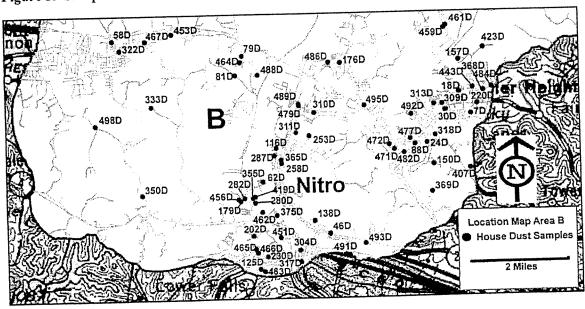


Figure 4. Sample stations for interior dust samples in Area B as shown in Fig. 2.

Tables

]	D	UTM X	ι	TM Y	Y		Date		
	4	426733	4	25275	0		19/2		
	5	437829	4	<u> 26043</u>	30		20/2		
	8	427854	4	25530)1		27/2		
	10	435593	4	26707	71	-	28/2		
	12	429224	4	2521	76		19/2		
	15	436098	4	2658	69			009	
	22	435356	4	2569	54			009	-
	27	419569	4	12507	93			009	-
	31	437885	4	12593	<u> 29</u>	_		2009	-
	32	427448	1	<u> 12542</u>	<u>52</u>			2009	-
	34	433394	4	<u> 12693</u>	<u>54</u>			2009	-
Γ	35	427711	١	<u> 42523</u>	41	5	/27/	2009	4
Γ	43	431110	1	42572	96	5	/28/	2009	4
	45	426029	L	<u>42583</u>	93	5	/21/	<u> 2009</u>	4
	49	423366		42587	20	5	/26/	<u> 2009</u>	4
Γ	50	437029		42625	92	5	/29/	<u> 2009</u>	4
Γ	55	422263		42579	29	5	/26/	<u> 2009</u>	4
	56	429859		42660)43	5	/27/	<u> 2009</u>	4
	58	429859		42602	294	5	/27/	2009	4
	61	429770		4253	775	15	5/19/	2009	
	67	425747		4262	017	<u> </u>	5/29/	<u> 2009</u>	
T	68	418672		4252	<u>504</u>	1	5/28	<u>/2009</u>	<u>니</u>
	70	419469		4255	311	:	5/26	<u>/2009</u>	
T	71	422089		4255	325		5/26	<u>/2009</u>	9
	73	437042		4258	001		5/20	/200	9
	74	425783		4258	022	2	5/21	/200	9
	76	422234		4255	903	3		/200	
T	79	424673	5	4259)46'	7		/200	
T	82	420451		4250	682	5	5/26	/200	9
ľ	86	41851	,	425	165	8	5/28	3/200	9
ľ	93	42935	1	4250	011	0)/20 0	
Ī	94	43031	3	426	049	6		7/200	
Ī	95	42003	4	425	814	6		6/200	
Ì	96	42815	6	425	964	3	5/2	7/200)9

Table 1. NAD83 UTM Zone 17N coordinates for soil samples taken along the right of way within the class area on the date specified.

										_	
I	D	U	TM X	U	TM	Y		Da		_	
9	97	4	31632	42	536	80	5/	20/2	200	9	
1	06	4	23854	42	2501	<u> 25</u>	_	28/2			
1	07	4	17755	42	2541	03	┼──	29/2		_	
1	09	4	27907	42	<u> 2495</u>	95		19/			
1	15	4	24259	42	<u> 2554</u>	136	+	27/		\neg	
1	17	4	129570	4:	<u> 2643</u>	394	+	/27/			
1	18*	4	133786	4	<u> 258</u> ()22	-	/20/			
	126	4	426559	4	<u> 260.</u>	301	+	/21/			
	127	<u> </u>	435971	4	<u> 257</u>	163		/20/			
	128		430121	4	262	<u>991</u>		/27/			
	129		422966	4	252	<u> 262</u>	-	/27			
	132		426979	4	256	097	_	/27			
	135		430066	4	261	162		5/27	<u>/20</u>	09	
	140		421035	1	1250	245	1	5/28	<u>/20</u>	09	
Γ	143	Τ	419760	1	1256	211		5/26	<u>/20</u>	09	
Γ	144	T	432092	4	<u> 1263</u>	360	_	5/28			
Γ	145	T	429126	1	4249	<u>533</u>	_	<u>5/19</u>			١
Γ	147	T	419479	L	4255	5725		5/26			١
	154	T	430179		4250	0770	5	5/19)/20	09	-
	157		419074		425	6 95 3	3	5/20			
Г	160	T	434804		426	<u>895</u> '	7	5/28			
	161	$ \top $	418353		425	<u>592</u>	0	5/2			_
T	162		419392		425	<u> 391</u>	0			009	
T	165		426252		426	146	5	5/2	1/2	009	
T	169		429592		425	356	0			009	
	170		432433	3	425	453	1	5/2	0/2	009	
	178		426388	3	425	852	28			009	_
T	180	$\neg \neg$	418388	3	425	5616	50			009	_
T	184		42265	5	425	5838	36			2009	_
Ī	188	1	41984	5	42	523	34			2009	
Ī	192		42887	3	42	<u>559′</u>	<u>78</u>			2009	_
Ī	200		43255		42	619	22			2009	-
Ì	201		43005	5	42	<u>575</u>	<u>96</u>			200	
	200	6	43244		42	574	<u>49</u>	5/	20/	200	9
1	* Appro	kima	te Coordinate	5							

Table 1 (cont.). NAD83 UTM Zone 17N coordinates for soil samples taken along the right of way within the class area on the date specified.

ID	UTM X	UTM Y	Date
209	432640	4265019	5/28/2009
210	424875	4258472	5/21/2009
216	434753	4263719	5/28/2009
218	424935	4258308	5/21/2009
220	427722	4259977	5/27/2009
223	434904	4262089	5/28/2009
227	435462	4259621	5/20/2009
228	430521	4267069	5/27/2009
248	420357	4257572	5/29/2009

Table 1 (cont.). NAD83 UTM Zone 17N coordinates for soil samples taken along the right of way within the class area on the date specified.

	, , , , , , , , , , , , , , , , , , ,			CITE!	DATE
ID	UTM X	UTM Y	ADDRESS	CITY	
107D	426898	4259016	242 Bills Creek Rd	Winfield	5/27/2009
116D	426148	4252631	1327 Park Ave	Nitro	5/29/2009
125D	425514	4248512	529 Fairview Dr	Saint Albans	6/3/2009
130D	425204	4258308	1371 Bills Creek Rd	Winfield	6/2/2009
138D	427446	4250167	206 Hillside Dr	Nitro	5/29/2009
150D	431541	4252008	19 Cedar View Dr	Cross Lanes	6/3/2009
151D	426161	4261898	221 Shawnee Estates	Winfield	5/20/2009
157D	432417	4255512	5403 Brookview Dr	Cross Lanes	6/3/2009
166D	424736	4258190	1680 Bills Creek Rd	Winfield	6/1/2009
176D	428425	4255478	434 Woodland Dr	Nitro	6/2/2009
	424890	4250855	1 Altamont Ave	Saint Albans	5/20/2009
179D	432420	4254437	5305 Linda Vista Dr	Cross Lanes	5/29/2009
18D		4249672	1195 Rosedale Dr	Saint Albans	6/3/2009
202D	425297	4254028	5293 Big Tyler Rd	Cross Lanes	5/21/2009
220D	433010	4248951	332 Birch St	Saint Albans	5/29/2009
230D	425773	 	163 Lake Shore Dr	Cross Lanes	5/20/2009
24D	431327	4252733	2192 N 21st St	Nitro	6/2/2009
253D	427312	4253027	133 Pine Dr	Poca	5/26/2009
256D	429275	4259502	603 6th St	Nitro	5/29/2009
258D	426324	4252121	314 Kentucky Ave	Saint Albans	5/21/2009
280D	425392	4250795	14 Orchard Ave	Saint Albans	6/3/2009
282D	425030	4250951	1123 W 11th St	Nitro	5/19/2009
287D	426082	4252371		Saint Albans	5/29/2009
304D	426913	4249168	106 5th Ave	Cross Lanes	5/20/2009
309D	431868	4254037	5234 Glow Dr	Cross Lanes	6/2/2009
30D	431964	4253830	5411 Karen Cir	Nitro	5/21/2009
310D	427498	4253812	3956 39th St E	Nitro	5/21/2009
311D	426854	4253139	2438 3rd Ave	Cross Lanes	5/20/2009
313D	431597	4254030	5337 Glow Dr	Saint Albans	5/27/200
317D	426938	4248770	911 Pennsylvania Ave Apt 1		5/20/200
318D	431613	4252979	5009 Frederick Dr	Cross Lanes	6/3/200
322D	420889	4255994	15-A Saunders St	Scott Depot	6/3/200
333D	421947	4254078	238 Joyce Rd	Scott Depot	5/20/200
33D	426176		208 Cannery Lane	Winfield	6/5/200
350D			460 Poplar Fork	Scott Depot	6/2/200
355D			208 Kentucky Ave	Saint Albans	0/2/200

Table 2. NAD83 UTM Zone 17N coordinates, street address, city, and collection date for interior dust samples taken in the Class Area.

-	UTM X	UTM Y	ADDRESS	CITY	DATE
		4252240	802 8th St	Nitro	5/19/2009
365D		4252240	5215 Swiss Dr	Cross Lanes	6/2/2009
368D			218 Majestic Dr	Cross Lanes	6/3/2009
369D		4251085	264 Oliver St	Saint Albans	5/27/2009
375D		4250357	4923 Dempsey Dr	Cross Lanes	5/20/2009
407D		4251860	1278 Poca River Rd	Poca	6/1/2009
415D		4257928	209 Virginia Ave	Saint Albans	5/21/2009
419D		4251001	5272 Claybank Rd	Cross Lanes	5/28/2009
423D	433233	4255907	160 Kilgore Rd	Scott Depot	6/5/2009
431D	421866	4258276	5258 Bailey Rd	Saint Albans	6/5/2009
443D	432865	4254571	120 Ranch Lake Blvd	Scott Depot	6/5/2009
445D	423046	4258590	228 Rust St	Saint Albans	5/21/2009
451D	426242	4249611		Poca	5/26/2009
452D	429234	4259662	155 Pine Dr	Scott Depot	5/26/2009
453D	422670	4256513	204 Willow Ridge	Winfield	5/27/2009
454D	426886	4259014	240 Bills Creek Rd	Saint Albans	5/27/2009
456D	424813	4250914	13 Flippin Ave	Poca	5/27/2009
457D	430502	4257578	15 Dairy Rd	Poca	5/28/2009
458D	429727	4258505	183 Silver St	Poca	5/28/2009
459D	431962	4256595	209 Country Rd	Poca	5/28/2009
460D	429681	4257222	152 Dairy Rd	Poca	5/28/2009
461D	432027	4256670	33-A Country Rd	Saint Albans	5/28/2009
462D	425631	4250478	114 Maryland Ave. N		5/28/2009
463D	425657	4248424	11 Clotine St	Saint Albans	6/1/2009
464D	424966	4255540	142 Poplar Dr	Scott Depot	6/1/200
465D	425374	4249219	14 Central Ave	Saint Albans	6/1/200
466D	425437	4249090	104 Central Ave	Saint Albans	
467D	421759	4256291	9 Meadow Wood	Scott Depot	6/1/200
468D	422675	4258485	171 Rocky Step Rd	Scott Depot	6/1/200
469D	422612	4258495	174 Rocky Step Rd	Scott Depot	6/1/200
46D	427967		727 1st Ave S	Nitro	5/19/200
471D	430218		17 Brick Lane	Cross Lanes	6/1/200
472D				Nitro	6/2/200
473D				Winfield	6/2/200
475D			73.1	Scott Depot	6/2/200
476D				Poca	6/2/200

Table 2 (cont.). NAD83 UTM Zone 17N coordinates, street address, city, and collection date for interior dust samples taken in the Class Area.

ID	UTM X	UTM Y	ADDRESS	CITY	DATE
477D	430768	4252880	170 Circle Dr	Cross Lanes	6/3/2009
477D	424628	4259766	251 Blue Lick Rd	Winfield	6/3/2009
479D	426967	4254110	4107 41st St	Nitro	6/4/2009
479D 481D	429889	4257616	83 Evergreen Circle	Poca	6/4/2009
481D 482D	430544	4252413	153 Brick Lane	Cross Lanes	6/4/2009
484D	433505	4254584	5315 Alpine Dr	Cross Lanes	6/5/2009
485D	426353	4261038	2046 Winfield Rd	Winfield	6/3/2009
	428025	4255495	106 Armour Rd	Nitro	6/4/2009
486D	430049	4257964	210 Spruce Lane	Poca	6/4/2009
487D	425564	4255107	108 Spruce Lane	Scott Depot	6/4/2009
488D		4254053	4011 40th St	Nitro	6/4/2009
489D	426972	4248998	121 Carson St	Saint Albans	6/4/2009
491D	428496	4253699	5337 Pioneer Dr	Cross Lanes	6/5/2009
492D	430814	4233033	721 Michigan Ave	Nitro	6/5/2009
493D	429158	4249373	219 Manilla Creek Rd	Nitro	6/4/2009
494D	430087	4254028	420-A Cross Lanes Rd	Nitro	6/4/2009
495D	429223	4254028	Route 1 Box 210-C	Poca	6/11/2009
496D	430723		Route 1 Box 218	Poca	6/11/2009
497D	430255	4262339	364 Poplar Fork	Scott Depot	6/11/2009
498D	420021	4253461	Route 1 Box 109-K	Poca	6/11/2009
499D	429798	4260193	Route 1 Box 123-C	Poca	6/12/2009
500D	431115	4261330	Route 1 Box 228	Poca	6/12/2009
503D	430256	4261244	706 Poinsetta Dr	Scott Depot	6/3/2009
58D	420641	4256323	111 Charleston St	Saint Albans	5/29/2009
62D	425665	4251502	146 Rolling Acres	Winfield	5/20/2009
78D	426672	4260312		Scott Depot	5/26/2009
79D	425031	4255750	117 Poplar Dr	Cross Lanes	6/5/2009
7D	432790	4253690	5120 Hopewell Dr	Scott Depot	6/5/2009
81D	424790		304 Scary Rd	Cross Lanes	6/4/2009
88D	430923	4252702	332 Goff Mountain Rd	CIU33 Editos	

Table 2 (cont.). NAD83 UTM Zone 17N coordinates, street address, city, and collection date for interior dust samples taken in the Class Area.

APPENDIX: Chain of Custody Forms

TENCON SOIL SAMPLE CHAIN OF CUSTODY

May 6. Malotte FOR LAB USE Initial for split sample receipt) |-|-(F) 360 Experse de 1 435 analyon by: 4435 439126 424734 ± 334 5/19/09 1:060 1:000 04: 4435 andlyon by 4435 5866 5866 FIELD NOTES and lyses by ソンディラ 15:021 5/14/04 3:18P Signature, Signature 487448 42542 \$ ± 19th 5/17/69 11.20A 429357 4250110 + 57+4 5/17/09 1232 A 5/19/01/01/72 2:30 TIME 5/19/09 DATE (Print Name) John Diamante 124: 4350776 + 284 429 592 425356 ± 1914 100 - Stolma 49824 \$5512 11944 Accuracy WITNESSED BY: (Print Name) W. JEFRREY CALENOER 426733 4252750 ACTUAL UTM Northing 430179 Ch. 1 LOCATION Easting Northing PLANNED UTM SAMPLED BY: Easting 54 45 50 23 2 ζ Site

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TENCON SOIL SAMPLE CHAIN OF CUSTODY

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TENCON SOIL SAMPLE CHAIN OF CUSTODY

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FOR LAB USE Initial for split sample receipt 1821/1813 By : 4435 SEM. FIELD NOTES TENCON SOIL SAMPLE CHAIN OF CUSTODY Signature 7 Signaturé 5/20/10 1640 5/20/A 1552 TIME DATE SAMPLED BY: (Print Name) LYNN & DAMLLY 191 WITNESSED BY: (Print Name) L. TEMPREY CANENDER 1425462 4259621 5.26 Northing | Accuracy ACTUAL UTM H37047 4758001 LOCATION Easting Northing PLANNED UTM Easting 33.7-Y 28 Site No.

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TENCON SOIL SAMPLE CHAIN OF CUSTODY	
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SAMPLED BY: (Print Name) JOHN DANNATE, JON Maidke Signature

WITNESSED BY: (Print Name)

Signature Signature

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FOR LAB USE analyze for EPA 4435 MISM for split sample receipt FIELD NOTES Signature _ 1.2:1 Signature. 11:04 5/2/6/ 11:58 12.3 5/29/00 1:08 5/29/04 11:33 TIME 5/29/09 5/29/09 124/29 DATE (Print Name) John Dig Mante 125 724 133 #/X 14 10 139 Accuracy 419845 4252384 420357 4257572 4250743 LASCOCK ACOLEH 417755 4254103 ACTUAL UTM WITNESSED BY: (Print Name) 5207+ YOUNG 419392 435310 Northing 419569 LOCATION Easting Northing PLANNED UTM Easting . SAMPLED BY: 1881 6.R 101 240 1600 10 00 Site No.

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TENCON DUST SAMPLE CHAIN OF CUSTODY

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TENCON DUST SAMPLE CHAIN OF CUSTODY

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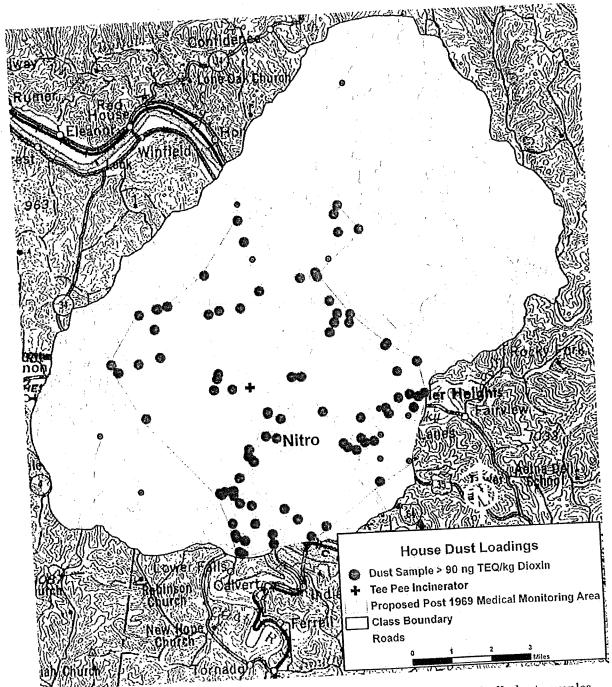


Figure 15. Proposed Post 1969 Area for Medical Monitoring based on bulk dust samples with dioxin loadings in excess of 90 ng TEQ/kg.

Figure 5.2

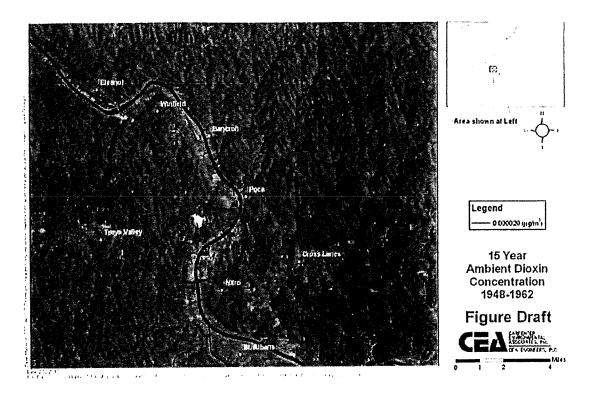


Figure 5.1

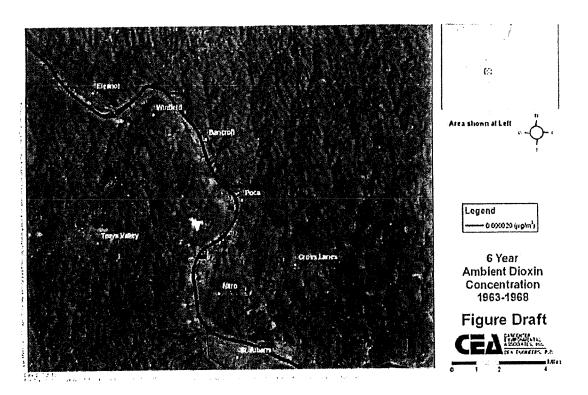
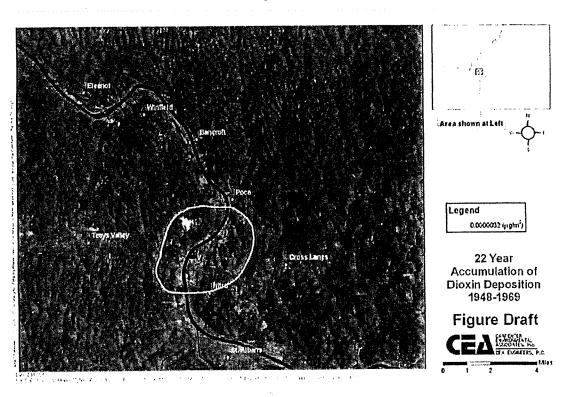
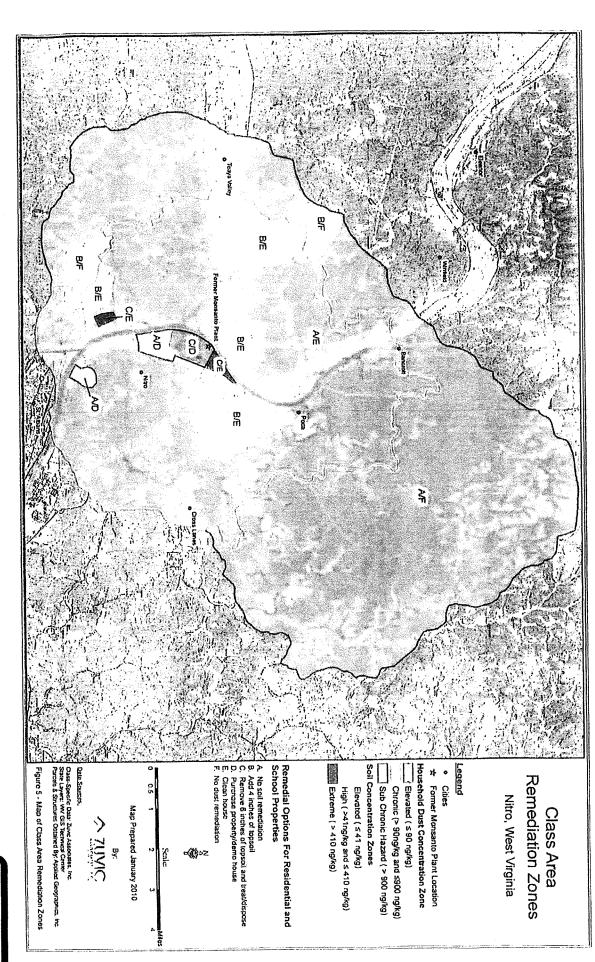


Figure 5.3





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